

# Alien Reference RNA QRT-PCR Detection Kit for Monitoring the Overall Performance of QRT-PCR Assays

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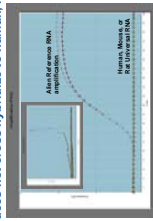
## Introduction

Exogenous RNA controls are increasingly used in QRT-PCR experiments to normalize variations that arise due to the presence of co-purified inhibitors (1,2). In addition, RNA controls allow assay standardization from experiment-to-experiment across laboratories, and the use of a highly sensitive and specific RNA control allows for the detection of common QRT-PCR inhibitors in RNA samples. When inhibitors are present, the Alien QRT-PCR Inhibitor Alert kit detects a delay in threshold cycle (Ct value) between Alien RNA amplified in the presence of sample RNA versus Alien RNA alone. The Inhibitor Alert kit employs SYBR Green chemistry, which provides high sensitivity, but does not allow multiplexing. We have recently expanded our line of exogenous controls to include TaqMan® primers and probe (FAM-and VIC-labeled) sets for detection of exogenous Alien RNA transcript in 1-step and 2-step QRT-PCR assays. As we will demonstrate, the Alien probe systems generate standard curves showing broad dynamic range (8-orders of magnitude) and single-copy detection of Alien RNA in 2-step QRT-PCR. In addition, the Alien probe system is sensitive to a variety of QRT-PCR inhibitors, and allow multiplex detection of Alien and sample RNA in the same tube.

## Results

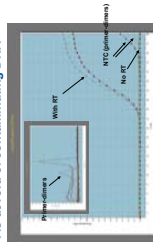
Alien Reference RNA template is a 500 bp *in vitro* transcript made from a computer generated sequence that possesses 5% GC-content and is non-homologous to sequences in GenBank.

### Part 1: Alien Reference RNA does not cross hybridize to human, mouse, or rat sequences



Amplification reactions contained 100nM of Alien RNA primers and either 10<sup>7</sup> copies of Alien RNA target or 50ng of Human, Mouse, or Rat Universal Reference RNA (Stratagene) in separate tubes. Amplifications were performed with Human, Mouse, or Rat Universal Reference RNA primers and Alien Reference RNA template. No amplification products were detected using Human, Mouse, or Rat Universal Reference RNA.

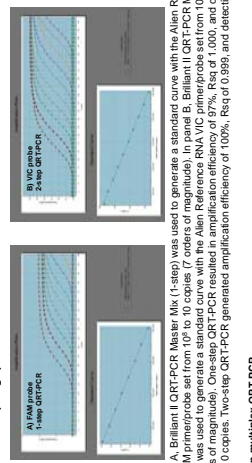
### Part 2: Alien Reference RNA is devoid of contaminating DNA



Amplification reactions were performed using Brilliant II SYBR Green QRT-PCR Master Mix (4-step), 10<sup>7</sup> copies of Alien Reference RNA, and 100nM of Alien RNA primers with and without the reverse transcriptase as indicated. No amplification is detected in the no-RT control.

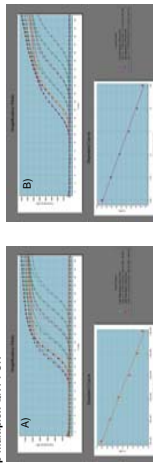
### Part 3: Alien Reference RNA primer/probe sets exhibit high amplification efficiency, sensitivity, and dynamic range

#### a) 1-step and 2-step singleplex QRT-PCR



In panel A, Brilliant II QRT-PCR Master Mix (1-step) was used to generate a standard curve with the Alien Reference RNA FAM primer/probe set from 10<sup>7</sup> to 10 copies (7 orders of magnitude). In panel B, Brilliant II QRT-PCR Master Mix (2-step) was used to generate a standard curve with the Alien Reference RNA VIC primer/probe set from 10<sup>7</sup> to 1 copy (8 orders of magnitude). One-step QRT-PCR resulted in amplification efficiency of 87%, Rq of 1.000, and detection limit of 10 copies. Two-step QRT-PCR generated amplification efficiency of 105%, Rq of 0.995, and detection limit of 1 copy.

#### b) 2-step multiplex QRT-PCR



Amplifications were performed using Brilliant II one-step QRT-PCR kit, 10<sup>7</sup> copies of Alien Reference RNA and 10ng Human Reference RNA were added to the reactions. Ethanol was titrated into the reactions at the concentrations indicated above. When 2.5% ethanol was added into the reaction, amplifications of Alien Reference RNA, Cyclophilin, B2M, and ADAM17 were inhibited. In contrast, amplification of Human Reference RNA was not inhibited. Human Reference RNA failed to amplify whereas all other targets produced delayed Ct's, suggesting that Alien Reference RNA is more sensitive to the inhibitory effects of ethanol compared to all the targets tested here.

## Results-continued

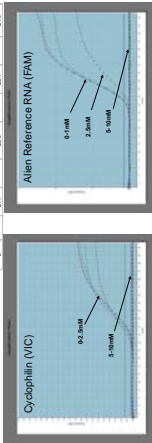
Brilliant Multiplex QPCR Master Mix was used to generate standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) individually or in multiplex. Amplifications were performed using Human QRT-PCR Reference RNA ranging from 10<sup>7</sup> to 10 copies. Panel A shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a singleplex reaction. Panel B shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel C shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel D shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel E shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel F shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel G shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel H shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel I shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel J shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel K shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel L shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel M shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel N shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel O shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel P shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel Q shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel R shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel S shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel T shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel U shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel V shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel W shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel X shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel Y shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction. Panel Z shows standard curves for cyclophilin (VIC) and Alien Reference RNA (FAM) in a multiplex reaction.

### Part 4: Alien Reference RNA amplification is sensitive to the presence of common QRT-PCR inhibitors

In order to detect inhibition, amplification of Alien Reference RNA alone (singleplex) is compared to the amplification of Alien Reference plus sample RNA (in singleplex or multiplex format). A delay in Ct of Alien Reference plus sample RNA suggests that the RNA sample contains inhibitors. To mimic this situation, we added two common QRT-PCR inhibitors (EDTA and ethanol) to the reactions. In addition, we added a 'no inhibitor' control. We demonstrate that Alien Reference RNA is sensitive to these known inhibitors in singleplex and multiplex probe formats, and hence should be able to detect a wide range of 'unknown' inhibitors in RNA samples.

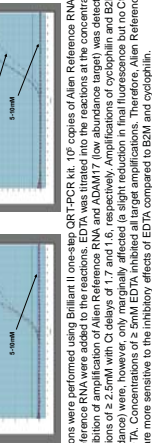
#### a) EDTA in one-step singleplex QRT-PCR:

EDTA (mM)	Human Reference RNA (Ct)	Cyclophilin (VIC) (Ct)	Alien Reference RNA (FAM) (Ct)
0	30.1	30.1	30.1
0.5	30.1	30.1	30.1
1.0	30.1	30.1	30.1
2.0	30.1	30.1	30.1
5.0	30.1	30.1	30.1
10.0	30.1	30.1	30.1
20.0	30.1	30.1	30.1
50.0	30.1	30.1	30.1
100.0	30.1	30.1	30.1



#### b) Ethanol in one-step singleplex QRT-PCR:

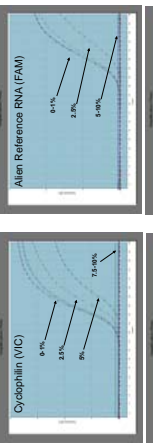
Ethanol (%)	Human Reference RNA (Ct)	Cyclophilin (VIC) (Ct)	Alien Reference RNA (FAM) (Ct)
0	30.1	30.1	30.1
0.5	30.1	30.1	30.1
1.0	30.1	30.1	30.1
2.0	30.1	30.1	30.1
5.0	30.1	30.1	30.1
10.0	30.1	30.1	30.1
20.0	30.1	30.1	30.1
50.0	30.1	30.1	30.1
100.0	30.1	30.1	30.1



Amplifications were performed using Brilliant II one-step QRT-PCR kit, 10<sup>7</sup> copies of Alien Reference RNA and 10ng Human Reference RNA were added to the reactions. EDTA was titrated into the reactions at the concentrations indicated above. Inhibition of amplification of Alien Reference RNA and ADAM17 (low abundance target) was detected at EDTA concentrations of 2.5mM with Ct delays of 1.7 and 1.6, respectively. Amplification of cyclophilin and B2M (medium to high abundance targets) was not inhibited at any of the tested concentrations. In contrast, amplification of ADAM17 was more sensitive to the inhibitory effects of EDTA compared to B2M and cyclophilin.

#### b) Ethanol in one-step singleplex QRT-PCR:

Ethanol (%)	Human Reference RNA (Ct)	Cyclophilin (VIC) (Ct)	Alien Reference RNA (FAM) (Ct)
0	30.1	30.1	30.1
0.5	30.1	30.1	30.1
1.0	30.1	30.1	30.1
2.0	30.1	30.1	30.1
5.0	30.1	30.1	30.1
10.0	30.1	30.1	30.1
20.0	30.1	30.1	30.1
50.0	30.1	30.1	30.1
100.0	30.1	30.1	30.1

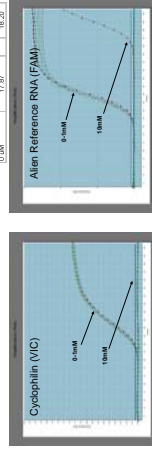


Amplifications were performed using Brilliant II one-step QRT-PCR kit, 10<sup>7</sup> copies of Alien Reference RNA and 10ng Human Reference RNA were added to the reactions. Ethanol was titrated into the reactions at the concentrations indicated above. When 2.5% ethanol was added into the reaction, amplifications of Alien Reference RNA, Cyclophilin, B2M, and ADAM17 were inhibited. In contrast, amplification of Human Reference RNA was not inhibited. Human Reference RNA failed to amplify whereas all other targets produced delayed Ct's, suggesting that Alien Reference RNA is more sensitive to the inhibitory effects of ethanol compared to all the targets tested here.

## Results-continued

### c) EDTA in two-step singleplex QRT-PCR:

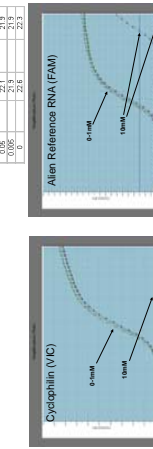
EDTA (mM)	Human Reference RNA (Ct)	Cyclophilin (VIC) (Ct)	Alien Reference RNA (FAM) (Ct)
0	30.1	30.1	30.1
0.5	30.1	30.1	30.1
1.0	30.1	30.1	30.1
2.0	30.1	30.1	30.1
5.0	30.1	30.1	30.1
10.0	30.1	30.1	30.1
20.0	30.1	30.1	30.1
50.0	30.1	30.1	30.1
100.0	30.1	30.1	30.1



Amplifications were performed using Brilliant II two-step QRT-PCR kit, 10<sup>7</sup> copies of Alien Reference RNA and 10ng Human Reference RNA were added to the cDNA synthesis reactions. EDTA was titrated into the reactions at the concentrations indicated above. Inhibition of amplification of both Alien Reference RNA and cyclophilin was observed at ≥ 10mM EDTA.

### d) EDTA in two-step multiplex QRT-PCR:

EDTA (mM)	Human Reference RNA (Ct)	Cyclophilin (VIC) (Ct)	Alien Reference RNA (FAM) (Ct)
0	30.1	30.1	30.1
0.5	30.1	30.1	30.1
1.0	30.1	30.1	30.1
2.0	30.1	30.1	30.1
5.0	30.1	30.1	30.1
10.0	30.1	30.1	30.1
20.0	30.1	30.1	30.1
50.0	30.1	30.1	30.1
100.0	30.1	30.1	30.1



Amplifications were performed using Brilliant Multiplex QPCR kit, 10<sup>7</sup> copies of Alien Reference RNA and 10ng Human Reference RNA were added to the cDNA synthesis reactions. EDTA was titrated into the reactions at the concentrations indicated above. Both Alien Reference RNA and cyclophilin amplifications are completely inhibited at 10mM EDTA similar to the results outlined above (part 4, section 0).

## Conclusions

Alien Reference RNA QRT-PCR detection kit is used for validating the quality of experimental RNA samples, interpreting the quality of QRT-PCR data, and monitoring the overall performance of QRT-PCR assay reagents and instrumentation.

- Alien Reference RNA is produced in large lots and subjected to stringent quality-control measures to ensure the availability of consistent reference RNA material over long-term experimental studies.

- The kit includes an exogenous RNA transcript, two primers and a TaqMan probe. The TaqMan probe is provided in with FAM (F300602) or VIC (F300605) dye for multiplexing flexibility.

- When multiplexing Alien Reference RNA with gene-of-interest (GOI), general multiplexing guidelines should be followed to minimize competition. These guidelines include using spectrally distinct fluorophores, a multiplex QPCR kit (e.g., Brilliant Multiplex-Kit), and titrating primer/probe concentration to higher abundance targets.

- The presence of an inhibitor in RNA samples is suspected; researchers are encouraged to re-purify or dilute the RNA sample (if it is not a no-inhibitor control). In addition, a two-step QRT-PCR format may be more appropriate to use due to increased tolerance of QRT-PCR inhibitors.

- Alien Reference RNA QRT-PCR detection kit will be commercially available on 11/26/08.

## References

- Smith, R.D. et al. Exogenous Reference RNA for Normalization of Real-time Quantitative PCR. *Bioinformatics*, 2006, 21: 3028-310.
- Nolan, T. et al. SPUD: A Quantitative PCR assay for the detection of inhibitors in nucleic acid preparations. *2008, Cellular Problems*, 2005, 19: p 54-58.
- Hartman, L.J. et al. Development of a novel internal positive control for TaqMan based assays. *Molecular and*

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